Review Article

Antioxidant and Anticancer Efficacy of Fruits

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Abstract

Fruits are good sources of natural antioxidants such as carotenoids, vitamins, phenols, flavonoids, dietary glutathionine, and endogenous metabolites. These antioxidants are capable of performing a number of functions including acting as free radical scavengers, peroxide decomposers, singlet and triplet oxygen quenchers, enzyme inhibitors and synergists. Active oxygen species are generated as by-products of normal metabolism. Increased levels of these active oxygen species or free radicals create oxidative stress which leads to avariety of biochemical and physiological injuries often resulting in impairment of metabolismand eventually, cell death. The consumption of fruits has been associated with low incidences and mortality rates of cancer (Ames et al., 1993; Dragsted et al., 1993; Willett, 1994) and heart disease (Verlangieri et al., 1985). Fruits also contain a wide range of flavonoids and phenolic acids. The main flavonoid sub groups in fruis are anthocyanins, proanthocyanins, flavonols, and catechins. Phenolic acids present in fruits are glycosides of hydroxylated derivatives of benzoic acid and cinnamic acid (Macheix et al., 1990). Good sources of natural antioxidants are found in many fruits, most notably prunes, raisins, blackberries, black currants, blueberries, raspberries, strawberries, grapes and pomegranates (Prior et al., 1998; Wang and Lin, 2000; Gil et al., 2000). The different antioxidant components found in fruits provide protection against harmful free radicals and have been associated with lower incidence and mortality rates of cancer and heart disease, in addition to a number of other health benefits. A number of researchers have identified the anticancer and antioxidant properties of different fruits using different methods. A comprehensive review of the work done and reported anticancer and antioxidant properties of different fruits has been done in this paper. This paper summarizes the traditional uses of some important fruit crops as well as their antioxidantand anticancer properties.

Keywords: Antioxidants; Jamun; Bael; Mulberry.

Introduction

The trend of agricultural production has deviated from mere focusing on maximizing production to optimizing the nutritional composition of fruits in order to promote health. In this regard, much reform in the agricultural practices has been put in place to enhance the nutritional content of the fruits. Fruits not only provide vitamins and minerals but also health promoting antioxidants. Antioxidants are the natural compounds that help in scavenging free radicals that are produced as by-products of normal metabolism. Interest in the role of antioxidants in human health has prompted research in the fields of horticulture and food science to assess fruit antioxidants, such as ascorbate, carotenoids, tocopherols, and phenolics. Fruits are good sources of both phenolics and ascorbate, and since fruits are often consumed fresh, antioxidant capacity is not lost due to any adverse effects of heat and oxidation during processing.

Cancer research has become a major area of scientific research supporting the foundations of modern biology to a great extent. Inspite of recent advances in cancer research and significant development in the field of synthetic drug chemistry, the global trend currently calls for discovery of new molecules of natural origin which are less toxic, endowed with fewer side effects and more potent in their mechanism of action. Regular consumption of fruits is associated with reduced risk of cancers and additive/ synergistic effects of phytochemicals in fruits are responsible for their potent antioxidant and anticancer activities. Natural products from some fruits offer new sources of drugs, but there are still a number of fruits in which all the active constituents have not yet been investigated although their medicinal effect is established by traditional use.

Carcinogens are becoming a big load on families and economies. Despite a good understanding of the molecular basis of the disease and advances in treatment, globally cancer is still a major cause of death. Estimates are that it will surpass cardiovascular disease as the leading cause of death, with higher incidence developing in countries that have minimal resources. Fruits consumed in our daily diet could be a solution to this burden by providing a chemoprotective and/or chemotherapeutic remedy. An estimated 12.6 million people are diagnosed with cancer every year and 7.6 million people die of the disease annually. Numerous lines of evidence suggest that lifestyle and nutrition play an important role in cancer development. As for nutrition, diets that are rich in fruits and vegetables have been shown to lower the risk of cancer incidence. In addition, several phytochemicals that are present in fruits and vegetables have been scientifically established to have properties that can prevent and treat this malevolent disease.

Antioxidant and Anticancer Efficacy of Fruits

It is increasingly being realized now that majority of the diseases are mainly due to the imbalance between pro-oxidant and anti-oxidant homeostasic phenomenon in the body. Pro-oxidant condition dominates either due to the increased generation of free radicals and/or their poor quenching/ scavenging in to the body. Free radicals are the fundamentals to any biochemical process, and represent an essential part of the aerobic life and our metabolism. They are continuously produced by the body, such as respiration and some cell mediated immune functions. There is a dynamic balance between the amount of free radicals generated in the body and anti-oxidants to quench and/or scavenge them and protect the body against the deleterious effects. Thus, the oxidant status in human reflects the dynamic balance between the anti-oxidant defense and pro-oxidant conditions and has been suggested as a useful tool in estimating the risk of oxidative damage (Khan, 2009). The anticancer and antioxidant properties reported in fruits has been summarized as under:

Olive (Olea europea; Oleaceae; Olive; Zaitoon)

Olive (*Olea europaea* L.) is an evergreen tree belonging to family Oleaceae. The tree has a native wild form *Olea cuspidate* L. which bears small fruits of pea size. Majority of the cultivated olive belongs to *Olea europaea* L. Olive oil is traditionally used in foods as it is considered good for health. It contains less saturated fatty acids as compared to other oils. Folkfore use olive oil for massaging to cure any kind of inflammation, allergy etc. Olive fruits also possess anti-oxidant properties. Presence of peroxisomes in olive (*Olea europaea* L.) fruits and different anti-oxidant enzymes in the plant tissue has been reported (Huertas and Rio, 2014).

Olive leaves are used for their antimicrobial (Bisignano et al., 1999; Pereira et al., 2007; Markin et al., 2003), gastroprotective (Dekanski et al., 2009), anti-oxidant (Benavente et al., 2000; Briante et al., 2003; Skrget et al., 2005; Somova et al., 2003), hypotensive (Khayyal et al., 2002), hypoglycaemic (Gonzalez et al., 1992; Al-Azzawie and Al-hamdani, 2006), antiarrhythmic (Somova et al., 2004), antiatherosclerotic (Wang et al., 2008), anti-viral (Lee-Huang et al., 2003; Micol et al., 2005), anti-tumor (Hamdi and Castellon, 2005; Abaza et al., 2007) and anti-inflammatory properties (Pieroni et al., 1996). Historically olive leaves have been used as a remedy for fever and other diseases such as malaria (Ciafardini and Zullo, 2002; Fernansez et al., 1999; Gucci et al., 1997). Essential oil of olea europaea L. leaves contain anti-inflammatory and alabgesic compounds like α -pinene, 2,6-dimethyloctane and 2-methoxy-3-isopropylpyrazine which show radical scavenging activity at different concentrations. Olea europaea L. essential oil at doses of 100, 200 and 300 mg/kg significantly reduced acetic acid induced abdominal constrictions and paw edema in male adult wistar rats and swiss albino mice (Haloni *et al.*, 2010).

Olive oil intake has been reported to prevent colon cancer. In particular, positive associations between elevated intakes of dietary fat and cancer of the colon (Armstrong and Doll, 1975), breast (La Vecchia et al., 1998), prostate (Chan et al., 1998) and ovary (Risch et al., 1994). Mediterranean diet has lower incidences of major illness such as cancer and cardiovascular disease, despite high intake of fat. The implication is that the health conferring benefits of the Mediterranean diet are mainly due to a higher consumption of fibre, fish, fruits and vegetables, but more recent research has focused on other important factors such as olives and olive oil (Owen et al., 2004). Phenols extracted from virgin olive oils on a series of in vitro systems that model important stages of colon carcinogenesis. Phenols extracted from virgin olive oil are capable of inhibiting several stages in colon carcinogenesis in vitro (Gill et al., 2005).

Bael (Aegle Marmelos; Rutaceae; Stone Apple; Bil)

Bael (*Aeglemarmelos* L.) is another Indian medicinal plant which has enormous traditional value against various diseases and many bioactive compounds have been isolated from this plant (Badam *et al.*, 2002; Gupta and Tondon, 2004). According to hindu mythology, the tree is another form of lord Kailashnath (Purohit and Vyas, 2004). Various phytoconstituents have been isolated from the various parts of *Aegle marmelos* L., which may be categorized as

Leaf

Skimmianine, Aegeline, Lupeol, Cineol, Citral, Citronella, Cuminaldehyde, Eugenol, Marmesinine

Bark

Skimmianine, Fagarine, Marmin

Fruit

Marmelosin, Luvangetin, Aurapten, Psoralen, Marmelide, Tannin

(Source: Maity et al., 2009)

Different parts of Bael are used for various therapeutic purposes such as for treatment of asthma, anaemia, Fractures, healing of wounds, swollen joints, high blood pressure, jaundice, diarrhea, healthy mind and brain typhoid troubles during pregnancy (Saswati, 2004). Aegle marmelos has been used as a herbal medicine for the management of diabetes mellitus in Ayurvedic, unani and siddha systems of medicine in India (Kar, 2003), Bangladesh (Lampronti, 2003), and Srilanka (Karinanayake, 1984). The unripe dries fruit is astringent, digestive, stomach ache and used to cure diarrhea and dysentery. Sweet drink prepared from the pulp of fruits produce soothing effect on the patients who have just recovered from bacillary dysentery. Anticancer potential of folk medicine used in Bangladesh as evaluated using extracts of Aegle marmelos for cytotoxic action using brine shrimp lethality assay, sea urchin eggs assay and MTT assay using tumor cell lines. The extracts showed toxicity on all used assays (Latica and Costa, 2005). Presence of skimmianine in the Aegle marmelos leaves contributes to the apoptosis activity (Gagetia et al.,2005).

Amla (Emblica Officinalis; Euphorbaceae; Gooseberry; Amla/Aonla)

Amla (Emblica officinalis L.) also called as amakali in Sanskrit has a special place in Ayurveda, which is the oldest health system in the world. Ayurveda appreciates and uses amla to treat a host of diseases and promote positive health. It is extensively used as a rejuvenatorin ayurveda. It is also used widely in combination with other two [chebulic and belleric] myrobalans [fruit bearing plant species] as triphala. Amla is the key ingredient in the popular ayurvedic recipe, chyavanaprasha. More than anything, it can be called as "King of Rasayana" [rejuvenation], owing to its multiple health benefits. The fruit is rich in quercetin, phyllaemblic compounds, gallic acid, tannins, flavanoids, pectin and vitamin C and also contain various polyphenolic compounds. A wide range of phytochemical components including terpenoids, alkaloids, flavonoids and tannins have been shown to possess useful biological activities (Kim et al., 2005; Arora et al., 2003). Many pharmacological studies have demonstrated the ability of the fruits to show anti-oxidant, anticarcinogenic, anti-tumor, anti-genotoxic, antiinflammatory activities, supporting its traditional use. Amla fruit contains more than 80% of water and nearly 20 times as much vitamin C as orange juice (Jain et al., 2000; Bharthakur et al., 1993).

The seeds yield a fixed oil (16%) which is brownish-yellow in colour. It contains fatty acids such as linoleic acid, linolenic acid, oleic acid, stearic acid, palmitic acid and myristic acid (Thakur *et al.*, 1989). The phytochemicals of this plant include hydrolysable tannins (Emblicanin A, Emblicanin B,

punigluconin, pedunculagin) (Ghosal *et al.*, 1996), flavonoids (Kaempferol 3 O apha L (6" methyl) rhamnopyranoside, Kaempferol 3 O alpha L (6" ethyl) amnopyranoside) (Rahman, 2007), alkaloids (Phyllantidine and phyllantine) (Khanna *et al.*, 1975). Phyllaemblic acid, a novel highly oxygenated norbisabolane is present in the roots of *P. emblica* (Zany *et al.*, 2003). Ellagic acid and lupeol are present in the roots of *P. emblica* (Kapoor, 1990; Rastogi *et al.*, 1993).

Because of the cooling nature, amla is a common ingredient in treatments for a burning sensation anywhere in the body and for many types of inflammations (Dang et al., 2011; Muthuraman et al., 2010; Nicolis et al., 2008) and fever; those are manifestations of pitta (fire) agitation (Williamson, 2000). Amla berry and other rasayanas are effective broad spectrum anti-oxidants and free radical scavengers, helping to reduce disease and slow the aging process. The use of amla as an antioxidant has been examined and authenticated (Kumaran et al., 2006; Rao et al., 2005; Naik et al., 2005; Bajpai et al., 2005; Bhattacharya et al., 2002; Anita et al., 2002; Scartezzini, 2000; Bandopathyay et al., 2000; Chaudhuri, 2003). Studies conducted in Japan have showed that amla is a potent scavenger of free radicals and amla preparations contained high levels of free-radical scavengers, super oxide dismutase (SOD) (Goshal, 1996). Aqueous fruit extracts of P. emblicasignificantly inhibited the growth of several human cancer cell lines at doses of 50-100 μg/ml (Ngamkitidechakul et al., 2010). Emblica officinalis polyphenols (EOP) are very effective in inducing apoptosis in human carcinoma cell lines (Rajeshkumar et al., 2003). Emblica officinalis has the potential to be useful in ameliorating the carcinogen-induced response in rat (Sarwat et al., 2008).

Valid research indicative of chemopreventive potential of *P.emblica* against skin carcinogenesis has also been reported (Garima et al., 2005). Amla fruit contains 18 compounds that inhibit the growth of tumor cells such as gastric and uterine cancer cells (Zhang et al., 2004). Chemoprevention with food phyto-chemicals is presently considered as one of the most important strategies to control cancer. Chemo-preventive potential of amla extract on 7,12dimethylbenz (a)anthracene (DMBA) induced skin tumorigenesis in Swiss albino mice have been found (Sancheti et al., 2005). Amla extract is potent in inhibiting *in vitro* cell proliferation towards human tumor cell lines, including human erythromyeloid K562, T-lymphoid Jurkat, B-lymphoid Raji, erythroleukemic HEL cell lines (Khan et al., 2002).

Jamun(Syzygium Cumini L.; Family; Black Plum; Jamun)

Jamun (Syzygium cumini L.) commonly known as black plum, Indian black berry, rose apple or 'jamun' is an important medicinal plant in various traditional systems of medicine. It as blackish-violet coloured juicy berry fruit that grows throughout the country particularly in rainy season. It is effective in the treatment of diabetes mellitus, inflammation, ulcers and diarrhea and preclinical studies have also shown it to possess chemo-preventive, radioprotective and antineoplastic properties. The plant is rich in compounds containing anthocyanins, glucosides, ellagic acid, isoquercetin, kermferol and myrecetin. The seeds are claimed to contain alkaloid, jambosine, and glycoside jambolin or antimellin, which halts the diastatic conversion of starch in to sugar (Swami et al. 2012). Jamun fruits are good source of iron, calcium, phosphorus, minerals, vitamin-C, sodium, potassium and carotene and are reported quite useful for the heart and liver dysfunction. The seeds of jamun are an effective home remedy commonly used by ayurvedic practitioners from times immemorial to control diabetes. It has been named as diabetic fighter as recommended by studies conducted by Indian Journal of Pharmacology, Division of Bhabha Atomic Research Center and Central Drug Research Institute - for its hypoglycaemic properties (lowering sugar level) (Baliga, 2011). Jamun seed has also gastro-protective properties. Jamun is effective in peptic ulcers, it helps to promote the mucosal defensive factors and antioxidant status and decrease lipid peroxidation. It has hypoglycaemia, anti-inflammatory, antibacterial and anti-HIV effects.

Mulberry (Morus Alba; Family; Mulberry; Shahtoot, Toot)

Many scientific and medical studies have examined mulberry fruit for its antioxidation and ant inflammation effects both in vitro and in vivo. Studies have reported on the chemical composition and nutritional potentials of some mulberry species worldwide (Ercisli and Orhan, 2007; Arabshahi-Delouee and Urooj, 2007; Darias-Martin et al., 2003; Elmacy and Altuo, 2002; Gao et al., 2000; Gerasopoulos and Stavroulakis, 1997). The deep coloured mulberry fruits are rich in phenolic compounds, including flavonoids, anthocyanins and carotenoids (Lin and Tang, 2007; Sass-Kiss, 2005; Zadernowski et al., 2005). Such natural substances extracted from plants have been shown to have greater antioxidant and anti-inflammatory effects and have been used for health maintenance and disease management since the beginning of recorded history (Krishnaswamy and Raghuramulu, 1998). Mulberry is traditionally used in Chinese medicines as a pharmaceutical for antifever diuretics, liver protection, eyesight improvement, blood pressure reduction and cardiovascular disease prevention. Dietary mulberry has been reported to have not only antioxidative, anti-inflammatory, antitumor and antidiabetic effects, but also cardio-vascular, hepatoand neuro-protective properties (Andallu et al., 2001; Huang et al., 2008; El-Beshbishy et al., 2006; Kimura et al., 2007; Isabelle et al., 2008; Wang et al., 2000). A study reported the proximate composition of mulberry fruits (Imran et al., 2010). Mulberry fruits contain 78.03-82.4 percent moisture (fresh weight). The ash content ranged between 0.46 and 0.87 g/ 100g dry weight (DW). The total lipid contents were in the range of 0.48-0.71 g/100g DW. The total protein contents of the fruits vary between 0.96 to 1.73 g/ 100g DW. The overall proximate composition of mulberry fruits indicates that it is a potential source of lipids, proteins, fibres, carbohydrates and hence, energy.

Among the phyto-chemicals in mulberry fruits, phenolic acids and flavonoids are regarded as major functional food components and are thought to contribute to the health effects of fruit derived products due to prevention of various diseases associated with oxidative stress (Lodovici et al., 2001). Many studies suggest flavonoids, a family of compounds with a C6-C3-C6 skeleton structure, display several biological activities, including antiallergic, antiviral, antitumor, and antiinflammatory action and antioxidant activity (Harborne and Williams, 2000). HPLC analysis showed that the main flavanols rich in mulberry fruit were rutin, morin, quercetin, and myricetin. These flavanols are reported to be effective antioxidants (Lu et al., 2006). Morin significantly reduced the tissue level of cyclosporine, a potent immunosuppressive agent with narrow therapeutic range, and dramatically decreased the nitric oxide production by the activated macrophages (Fang et al., 2005). In addition, C3G isolated from mulberry fruits inhibited the cerebral ischemic damage caused by oxygen glucose deprivation in PC12 cells (Kang et al., 2006). Black mulberry fruit juice inhibited the human cytochrome CYP3A activity in the pooled human liver microsome system (Kim et al., 2006).

Mulberry anthocyanins, cyaniding 3-rutinoside, and cyanidin 3-glucoside, exhibited an inhibitory effect on the migration and invasion of a human lung cancer cell line by regulating the activation of c-Jun and NF-kB (Chen *et al.*, 2006). Mulberry fruit extracts

result in human glioma cell death in vitro through the reactive oxygen species (ROS)-dependent mitochondrial pathway and glioma tumor growth in vivo via reduction of tumor cell proliferation and the induction of apoptosis (Jeong et al., 2010). Mulberry fruit colour does not affect the antioxidant capacity. Researches made on pink, white and black fruit exhibited no significant effect on the antioxidant potential of the mulberry fruit. However, location has a role to play in the antioxidant capacity of mulberry fruit (Bajpai et al., 2014). The source from genotypes from different sampling locations is more important as compared to fruit colour in determining antioxidant capacity, total polyphenol content (TPC), total flavonoid content (TFC) and total anthocyanin content (TAC) in M. alba fruit.

Ber (Ziziphus Mauritiana L.; Rhamnaceae; Chinese Apple; Ber)

Ber (Ziziphus mauritiana L.) also known as Chinese apple, Jujube, Indian plum and Masau is a tropical fruit tree species belonging to the family Rhamaceae. Ber, though an underutilized fruit, has medicinal properties as well. The fruits are applied on cuts and ulcers, are employed in pulmonary ailments and fevers; and mixed with salt and chilli peppers, are given in indigestion and biliousness. The dried ripe fruit is a mild laxative. The seeds are sedative and are taken, sometimes with butter milk, to halt nausea, vomiting and abdominal pains in pregnancy. They check diarrhoea and are poultice on wounds. Mixed with oil, they are rubbed on rheumatic areas. The leaves are applied as poultices and are helpful in liver troubles, asthma and fever and, together with catechu, are administered when an astringent is needed, as on wounds. The bitter astringent bark decoction is taken to halt diarrhea and dysentery and relieve gingivitis. An infusion of flowers serves as an eye lotion.

Karonda (Carissa Carandas; Apocynaceae; Karanda; Karonda)

Karonda (*Carissa carandas*) another underutilized tropical fruit of Indian subcontinent. The karonda fruit is an astringent, antiscorbutic and used as a remedy for biliousness and for cure of anaemia. In traditional medicine, the fruit is used to improve female libido and to remove worms from the intestinal tract. The fruit has anti-microbial and anti-fungal properties and its juice used to clean old wounds which have become infected. The fruit have an analgesic action as well as an anti-inflammatory one.

The juice can be applied to the skin to relieve any skin problems. Traditionally karonda has been used to treat anorexia and insanity. A leaf decoction of karonda is used against fever, diarrhea and earache. The roots serve as a stomachic, vermifuge, remedy for itches and insect repellant. Traditional healers of Chhattisgarh having expertise in treatment of different types of cancer from karonda. They use its different plant parts to dress the cancerous wounds and to kill the maggots.

Phalsa (*Grewia Subinequalis* L.;Tilliaceae; The Elusive Berry; Phalsa)

Phalsa (Grewia subinequalis L.) is indigenous throughout the Himalayan region. Fruits are globose drupe, 1.8-2.2 cm in diameter, indistinctly lobed, red or purple, finely warty and with stellate hairs and ripe fruits are eaten as a dessert. Their taste and flavor are very much liked and they fetch high prices. According to Ayurveda, the ancient Indian treatise on medicine, the fruits are cooling, tonic and aphrodisiac, they allay thirst and burning sensation, remove biliousness, cure inflammation, heart and blood disorders and fevers. The bark is used as a demulcent. It cures urinary troubles and relieves burning in the vagina. Fruit extracts of phalsa exhibit antitumor and in-vitro cytotoxic activity against ehrlich's ascites carcinoma cell lines (Kakotiet al., 2011).

Pomegranate(*Punica Granatum* L.; Punicaceae; Pomegranate; Anar)

Pomegranate (Punica granatum L.) fruit is also a potential source of antioxidants and exhibits anticancer activity. The peels (pericarp, rind or hull) amounts to approximately 60% of the pomegranate fruit weight (Lansky and Newman, 2007). Pomegranate fruit extracts are rich in ellagitannins (ETs) that act as antioxidant and anticancer agents especially against breast and colon cancer (Lansky and Newman, 2007; Adams et al., 2010; Kasimsetty et al., 2010; Sharma et al., 2010). Pomegranate peel extracts have exhibited marked antioxidant capacity in several studies using solvents such as methanol and a mixture of methanol, acetone, ethyl acetate and water (Ghasemian et al., 2006; Negi et al., 2002; Panichayupakarananta et al., 2010). Powdered peels of pomegranate also exhibit antioxidant capacity (Hasnaouiet al., 2014). Ellagitannins (ETs) are the predominant phenolics in pomegranate peel, mesocarp and arils extracts (Fischer et al., 2011). Upon consumption, these ETs are hydrolysed releasing EA, which is then converted to urolithin derivatives by gut microflora (Seeram et al., 2007). EA is known for its antioxidant, anti-inflammatory and anticarcinogenic properties, besides it is considered as a biomarker for human bioavailability studies involving consumption of ETs containing food, as it is detected in human plasma (Seeram et al., 2004). The 50% ethanolic extract of the peels standardized to contain the highest percent of EA (1.9 \pm 0.1%) exhibited a pronounced antioxidant activity $(IC50=0.50\pm0.9 \text{ mg/ml})$ compared to the fruit $(0.3\pm0.9 \text{ mg/ml})$ 0.05% EA) and pulp (0.02± 0.01 % EA) extracts showing an IC50 of $1.09 \pm 0.5 \text{ mg/ml}$ and 13.6 ± 0.8 mg/ml, respectively. These results show a direct correlation between the EA content in pomegranate extracts and its ability in quenching free radicals. The contents of total phenolics in pomegranate peel extract was reported to be 10-fold as much as its content in the pulp extract, which causes its stronger antioxidant ability (Li et al., 2006)

The anticancer activity of pomegranate fruit, peels and pulp extracts has been standardized to their EA content (Motaal and Sherif, 2011). Cytotoxicity tested against MCF-7 cell line (human breast carcinoma) and HCT-116 cell line (human colon carcinoma) along with doxorubicin as a positive control. The anticancer activity against both the cell lines increased by increasing the extract concentrations from 1 to 10µg/ml for all three extracts. The pomegranate peel extract showed a pronounced cytotoxic activity against MCF-7 cells compared to the other two extracts at a concentration of 10µg/ml, confirmed by the least IC50 (7.7 \pm 0.01 μ g/ml). It has been reported that the ellagitannins and their intestinal bacterial metabolites, urolithins, released in the colon upon consumption of pomegranate juice could potentially diminish the risk of colon cancer development, by inhibiting cell proliferation and inducing apoptosis (Kasimsetty et al., 2010). Also pomegranate ET-derived compounds especially urolithin B have potential for the prevention of estrogen-responsive breast cancers by significantly inhibiting testosterone-induced MCF-7 cell proliferation (Adams et al., 2010).

Conclusion

It is quite evident from this review that fruits are a potential source of phytoconstituents that reveals their usefulness in diet of humans. Fruits as well as their parts can be utilized in various ways for the treatment of various disorders in human beings such as diabetes, liver toxicity, fungal infections, microbial infections, inflammations etc. These are potential

source of antioxidants that can help scavenge harmful free radicals produced inside the body. Their possible role in cancer prevention has been researched and professed. Still, still there is a need to explore the medicinal properties of various fruits in order to utilize these in other therapeutic activities.

References

- Abaza, L., Talorete, T. P., Yamada, P., Kurita, Y., Zarrouk, M and Isoda, H. Induction of growth inhibition and differentiation of human leukemia HL-60 cells by a Tunisian gerboui olive leaf extract. Biosci. Biotechnol. Biochem. 2007; 71:1306-1312.
- Adams, L. S.; Zhang, Y.; Seeram, N. P.; Heber, D. and Chen, S. Pomegranate ellagitannin-derived compounds exhibit antiproliferative and antiaromatase activity in breast cancer cells in vitro. Cancer Prevention Research (Phila Pa) 2010; 3(1):108-13.
- Al-Azzawie, H and Alhamdani, M.S.S. Hypoglycemic and antioxidant effect of oleuropein in alloxan-diabetic rabbits. Life Science 2006; 78:1371-1377.
- 4. Ames, B. M.; Shigena, M. K. and Hagen, T. M. Oxidants, antioxidants and the degenerative diseases of aging. Proc. Natl. Acad. Sci. USA. 1993; 90: 7915-7922.
- Andallu, B., Suryakantham, V., Lakshmi Srikanthi, B. and Reddy, G. K. Effect of mulberry (Morus indica L.) therapy on plasma and erythrocyte membrane lipids in patients with type 2 diabetes. Clinical Chim Acta. 2001; 314:47-53.
- Anila L and Vijayalakshmi NR. Flavonoids from Emblica officinalis and Mangifera indica effectiveness for dyslipidemia. Journal of Ethnopharmacology 2002; 79(1):81-87.
- 7. Arabshahi-Delouee, S. and Urooj, A. Antioxidant properties of various solvent extracts of mulberry (Morus indica L.) leaves. Food Chemistry. 2007; 102:1233-40.
- Armstrong BK, Doll R. Environmental factors and the incidence and mortality from cancer in different countries with special reference to dietary practice. Interntional Journal of Cancer 1975; 15: 617–631.
- 9. Arora, S., Kaur, K. and Kaur, S. Indian medicinal plants as a reservoir of protective phytochemicals. Teratog Carcinog Mutagen. (suppl1): 2003.p.295-300.
- 10. Badam L, Bedekar S.S., Sonawane K.B. and Joshi S.P. "Invitro antiviral activity of Bael(Aegle marmelos Corr.) upon human Cox sackiviruses B1-B6. Journal of Communicable Diseases 2002; 34:88.
- 11. Bajpai, P. K.; Warghat, A. R.; Dhar, P.; Kant, A.; Srivastava, R. B. and Stobdan, T. Variability and relationship of fruit colour and sampling location

- with antioxidant capacities and bioactive content in Morus alba L. fruit from Trans-Himalaya, India. Food Science and Technology. 2014; 59(2):981-988.
- 12. Bajpai M, Pande A, Tewari SK, Prakash D. 2005. Phenolic contents and antioxidant activity of some food and medicinal plants. International Journal of Food Science Nutrition 2005; 56(4):287-291.
- 13. Baliga, M.S. Anticancer, chemopreventive and radioprotective potential of black plum (Eugenia jambolana L.). Asian Pacific Journal of Cancer Prevention. 2011; 12(1):3-15.
- 14. Bandyopadhyay SK, Pakrashi SC, Pakrashi A. The role of antioxidant activity of Phyllanthus emblica fruits on prevention from indomethacin induced gastric ulcer. Journal of Ethnopharmacology 200; 70(2):171-176.
- Benavente-Garcia, O., Castillo, J., Lorente, J., Ortuno, A and Del Rio, J. A. Antioxidant activity of phenolics extracted from Oleaeuropaea L. leaves. Food Chemistry 2000; 68:457-462.
- 16. Bharthakur, N.N. and Arnold, N.P. Chemical analysis of the emblic (Phyllanthus emblica L) and its potential as a good source. Scientia Horticulture 1991; 47:99-105.
- 17. Bhattacharya SK, Bhattacharya A, Sairam K, Ghosal S. Effect of bioactive tannoid principles of Emblica officinalis on ischemiareperfusion- induced oxidative stress in rat heart. Phytomedicine 2002; 9(2): 171-174.
- 18. Bisignano, G., Tomaino, A., Cascio, R.L., Crisafi, G., Uccella, N. and Saija, A. On the in vitro antimicrobial activity of oleuropein and hydroxytyrosol. Journal of Pharmacy and Pharmacology 1999; 51:971-974.
- 19. Briante, R., Febbraio, F and Nucci, R. Antioxidant properties of low molecular weight phenols present in the Mediterranean diet. Journal of Agriculture and Food Chemistry 2003; 51:6975-6981.
- 20. Chan, JM., Stampfer, MJ and Giovannucci, EL. What causes prostate cancer? A brief summary of the epidemiology. Semin Cancer Biology 1998; 8: 263–273.
- Chaudhuri, R. K., Guttierez, G. and Mustafa, S. Low Molecular-Weight Tannins of Phyllanthus emblica: A New Class of Anti- Aging Ingredients. Proceedings Active Ingredients Conference, Paris. 2003.
- 22. Chen, P.N., Chu, S.C., Chiou, H.L., Kuo, W.H., Chiang, C.L. and Hsieh, Y.S. Mulberry anthocyanins, cyaniding 3-rutinoside and cyaniding 3-glucoside, exhibited an inhibitory effect on the migration and invasion of a human lung cancer cell line. Cancer Lett. 2006; 235:248-59.
- 23. Ciafardini, G and Zullo, B. A. Microbiological activity in stored olive oil. International Journal of Food Microbiology 2002; 75:111-118.

- 24. Dang, G.K., Parekar, R.R., Kamat, S.K., Scindia, A.M. and Rege, N.N. Antiinflammatory activity of Phyllanthus emblica, Plumbago zeylanica and Cyperus rotundus in acute models of inflammation. Phytother Res, 2011; 25:904-908.
- Darias-Martin, J., Lobo-Rodrigo, G., Hernandez-Cordero, J., Diaz-Diaz, E. and Diaz-Romero, C. Alcoholic beverages obtained from black mulberry. Food Technology Biotechnology 2003; 41:173-6.
- Dekanski, D., Janicijevic-Hudomal, S., Tadic, V., Markovic, G., Arsic, I. and Mestrovic, D.M. Photochemical analysis and gastroprotective activity of an olive leaf extract. J. Serb. Chem. Soc. 2009; 74:367-377.
- Dragsted, L.O., Strube, M. and Larsen, J.C. Cancerprotective factors in fruits and vegetables: biochemical and biological background. Pharmacology and Toxicology 1993; 72 (Suppl. 1): 116-135.
- 28. El-Beshbishy, H.A.; Singab, A. N.; Sinkkonen, J. and Philaja, K. Hypolipidemic and antioxidant effects of Morus alba L. (Egyptian mulberry) root bark fractions supplementation in cholesterol-fed rats. Life Science 2006; 78:2724-33.
- 29. Elmacy, Y. and Altuo, T. Flavor evaluation of three black mulberry (Morus nigra) cultivars using GC-MS, chemical and sensory data. Journal of Science and Food Agriculture 2002; 82:632-5.
- Ercisli, S. and Orhan, E. Chemical composition of white (Morus alba), red (Morus alba) and black (M. nigra) mulberry fruits. Food Chemistry 2007; 103:1380-4.
- 31. Fang, S.H.; Hou, Y.C. and Chao, P.D. Pharmacokinetic and pharmacodynamic interactions of morin and cyclosporine. Toxicology and Applied Pharmacology 2005; 205:65-70.
- 32. Fernandez-Escobar, R., Moreno, R and Garcia-Creus, M. Seasonal changes of mineral nutrients in olive leaves during the alternate-bearing cycle. Scientia Horticulture 1999; 82:25-45.
- 33. Fischer, U.A.; Carle, R. and Kammerer, D.R. Identification and quantification of phenolic compounds from pomegranate (Punica granatum L.) peel, mesocarp, aril, and differently produced juices by HPLC-DAD-ESI/MSn. Food Chemistry 2011; 127:807-21.
- 34. Gagetia, G.C., Venkatesh, P. and Baliga, M.S. "Aegle marmelos (L.) Correa inhibits the proliferation of transplanted Ehrlich ascites carcinoma in mice", Biology and Pharma Bulletin 2005; 28:58-64.
- 35. Gao, X.; Ohlander, M.; Jeppsson, N.; Bjork, L. and Trajkovski, V. Changes in antioxidant effects and their relationship to phytonutrients in fruits of sea buckthorn (Hippophae rhamnoides L.) during maturation. Journal Agriculture Food Chemistry 2000; 48:1485-90.

- 36. Garima, S., Jindal, A., Goyal, R.K. Chemopreventive Action of Emblica officinalis on Skin Carcinogenesis in mice. Asian Pacific Journal of Cancer Prevention 2005; 6:197 201.
- 37. Gerasopoulos, D. and Stavroulakis, G. Quality characteristics of four mulberry (Morus spp.) cultivars in the area of Chania, Greece. Journal Science and Food Agriculture 1997; 73:261-4.
- 38. Ghasemian, A.; Mehrabian, S. and Majid, A. Peel extracts of two Iranian cultivars of pomegranate (Punica granatum L.) have antioxidant and antimutagenic activities. Pakistan Journal of Biological Science 2006;9(7):1402-5.
- 39. Ghosal, S., Tripathi, V.K. and Chauhan, S. Active constituent of Emblicaofficinalis: part 1st the chemistry and antioxidant effects of two new hydrolysable tannins, emblicanin A and B. Indian Journal of Chemistry 1996; 35:941–948.
- 40. Gil, M.I.; Tomas-Barberan, F. A.; Hess-Pierce, B.; Holocroft, D.M. and Kader, A.A. Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. Journal Agriculture and Food Chemistry 2000; 48:4581-4589.
- 41. Gill, C.I.R., Boyd, A., McDermott, E., McCann, M., Servili, M., Selvaggini, R., Taticchi, A., Esposto, S., Montedoro, G., McGlynn, H. and Rowland, I. Potential anti-cancer effects of virgin olive oil phenols on colorectal carcinogenesis models in vitro. International Journal of Cancer 2005; 117:1-7.
- 42. Gonzalez, M., Zarzuelo, A., Gamez, M.J., Utrilla, M.P., Jimenez, J and Osuna, I. 1992. Hypoglycemic activity of olive leaf. Planta Med. 1992; 58:513-515.
- 43. Gucci, R., Lombardini, L and Tattini, M. Analysis of leaf water relations in leaves of two olive (Olea europaea) cultivars differing in tolerance to salinity. Tree Physiology 1997; 17:13-21.
- 44. Gupta, A.K. and Tondon N. "Review on Indian medicinal plants", Indian council of medicinal research, New Delhi, 2004; 312.
- 45. Hamdi, H. K and Castellon, R. Oleuropein, a nontoxic olive iridoid, is an anti-tumor agent and cytoskeleton disruptor. Biochem. Biophys. Res. Commun. 2005; 334:769-778.
- 46. Harborne, J.B. and Williams, C.A. Advances in flavanoid research since 1992. Phytochemistry 2000; 55:481-504.
- 47. Hasnaoui, N.; Wathelet, B. and Araujo, A.J. Valorization of pomegranate peel from 12 cultivars: Dietary fibre composition, antioxidant capacity and functional properties. Food Chemistry 2014; 160:196-203.
- 48. Heinonen, I. M.; Meyer, A. S. and Frankel, E. N. Antioxidant activity of berry phenolics on human low-density lipoprotein and liposome oxidation. Journal Agric. Food Chemistry 1998; 46: 4107-4112.
- 49. Huang, H. P.; Shih, Y.W.; Chang, Y. C.; Hung, C. N.

- and Wang, C.J. Chemoinhibitory effect of mulberry anthocyanins on melanoma metastasis involved in the Ras/PI3K pathway. Journal Agriculture and Food Chemistry 2008; 56:9286-93.
- Imran, M.; Khan, H.; Shah, M.; Khan, R. and Khan, F. Chemical composition and antioxidant activity of certain Morus species. Journal Zhejiang University Science B. 2010; 11:973-80.
- Isabelle, M.; Lee, B. L.; Ong, C. N.; Liu, X. and Huang, D. 2008. Peroxyl radical scavenging capacity, polyphenolics, and lipophilic antioxidant profiles of mulberry fruits cultivated in southern China. Journal of Agriculture and Food Chemistry 2008; 56:9410-6.
- 52. Jain, S.K. and Akhurdiya, D.S. Anola: Potential fruit for processing Delhi Garden Mega 2000; 38:50-51.
- 53. Jeong, J.C.; Jang, S.W.; Kim, T.H.; Kwon, C. H. and Kim, Y.K. Mulberry fruit (Moris fructus) extracts induce human glioma cell death in vitro through ROS-dependent mitochondrial pathway and inhibits glioma tumor growth in vivo.Nutr Cancer. 2010; 62:402-12.
- 54. Kakoti, B.B.; Selvan, V.T.; Manikandan, L.; Gupta, M.; Mazumder, U.K. and Das, B. Antitumor and invitro cytotoxic activity of Grewia asiatica L. against ehrlich's ascites carcinoma cell lines. Pharmacologyonline. 2011; 3(3):956-960.
- 55. Kalt, W.; Prange, R.K. and Lidster, P.D. Postharvest color development of strawberries: Influence of maturity, temperature and light. Canadian Journal of Plant Science. 1993; 73: 541-548.
- Kang, T.H.; Hur, J.Y.; Kim, H.B.; Ryu, J.H. and Kim, S.Y. Neuroprotective effects of the cyaniding-3-Obeta-d-glucopyranoside isolated from mulberry fruit against cerebral ischemia. Neurosci Lett. 2006; 391:122-6.
- 57. Kapoor, L.D. Handbook of Ayurvedic Medicinal Plants. CRC Press. Boca taton. 1990.
- 58. Kar, A., Choudhry, B.K. and Bandhopadhyay, N.G. "Comparative evaluation of hypoglycemic activity of some Indian medicinal plants in alloxan diabetic rats". J Ethnopharmacol. 2003; 84:105-108.
- 59. Karunanayake, E.H., Welihinda, J., Sirimanne, S.R. and Sinnadorai, G. "Oral hypoglycemic activity of some medicinal plants of Sri Lanka". J Ethnopharmacol, 1984; 11:223-231.
- Kasimsetty, S. G.; Bialonska, D.; Reddy, M. K.; Ma, G.; Khan, S. I. and Ferreira, D. Colon cancer chemopreventive activities of pomegranate ellagitannins and urolithins. Journal of Agricultural Food Chemistry. 2010; 58(4):2180-7.
- 61. Khan, M. T.; Lampronti, I.; Martello, D.; Bianchi, N.; Jabbar, S.; Choudhuri, M. S.; Datta, B. K. and Gambari, R. Identification of pyrogallol as an antiproliferative compound present in extracts from the medicinal plant Emblica officinalis: Effects on in

- vitro cell growth of human tumor cell lines. International Journal of Oncology. 2002; 21(1):187-192.
- 62. Khayyal, M.T., El-Ghazaly, M.A., Abdallah, D.M., Nassar, N. N., Okpanyi, S. N and Kreuter, M. H. Blood pressure lowering effect of an olive leaf extract (Olea europaea) in L-NAME induced hypertension in rats. Arzneimittel-Forsch. 2002; 52:797-802.
- Kim, H.; Yoon, Y.J.; Shon, J.H.; Cha, I.J.; Shin, J.G. and Liu, K.H. Inhibitory effects of fruit juices on CYP3A activity. Drug Metab Dispos. 2006; 34:521-3.
- 64. Kim, H.J., Yokozawat, K., Tohda, C., Rao, T.P and Juneja, L.R. Influence of Amla (Emblica Officinalis Gaertnl) on hypercholesterolemia and lipid peroxidation in cholesterol-fed rats. J Nutr Sci Vitaminol, 2005; 51:413-418.
- 65. Kimura, T.; Nakagawa, K.; Kunota, H.; Kojima, Y.; Goto, Y. and Yamagishi, K. Food-grade mulberry powder enriched with 1-deoxynojirimycin suppresses the elevation of postprandial blood glucose in humans. Journal Agric. Food Chem. 2007; 55:5869-74.
- Krishnaswamy, K. and Raghuramulu, N. 1998. Bioactive phytochemicals with emphasis on dietary practices. Indian Journal of Medical Research. 1998; 108:167-81.
- 67. Kumaran, A. and Karunakaran, R. J. 2006. Nitric oxide radical scavenging active components from Phyllanthus emblica L. Plant Foods Hum Nutr. 2006; 61(1):1-5.
- La Vecchia, C., Favero, A and Franceschi, S. Monounsatuated and other types of fat, and the risk of breast cancer. Eur J Cancer Prev 1998; 7:461–464.
- Lampronti, I., Martello, D., Bianchi, N., Borgatti, M., Lambrtini, E., Piva, R., Jabbars, S., Choudhuri, M.S., Khan, M.T. and Gambari, R. "In Vitro antiproliferative effect on human tumor cell lines of extracts from the bangladesi medicinal plant Aegle marmelos Correa." Phytomedicine, 2003; 10: 300-308.
- Lansky, E. P. and Newman, R.A. Punica granatum (pomegranate) and its potential for prevention and treatment of inflammation and cancer. Journal of Ethnopharmacology. 2007; 109(2): 177-206.
- 71. Latica, V. and Costa, L. Evaluation of anticancer potential used in Bangladeshi folk medicine. J. Ethnopharmacol., 2005; 99:21-38.
- 72. Lee-Huang, S., Zhang, L., Huang, P. L and Chang, Y. T. Anti-HIV activity of olive leaf extract (OLE) and modulation of host cell gene expression by HIV-1 infection and OLE treatment. Biochem.Biophys. Res. Commun. 2003; 307:1029-1037.
- 73. Li, Y,; Guo, C.; Yang, J.; Wei, J.; Xu, J. and Cheng, S. Evaluation of antioxidant properties of pomegranate peel extract in comparison with

- pomegranate pulp extract. Food Chemistry. 2006; 96(2):254-60.
- 74. Lin, J.Y. and Tang, C.Y. Determination of total phenolic and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation. Food Chemistry. 2007; 101:140-7.
- 75. Lodovici, M.; Guglielmi, F.; Meoni, M. and Dolara, P. Effect of natural phenolic acids on DNA oxidation in vitro. Food Chem Toxicol. 2001; 39:1205-10.
- 76. Lu Xi, W. L.; Wei, H.; Yang, Z. and Wei, W. Structureactivity relationship of flavonoids in antioxidant activity. Food Science. 2006; 27:233-7.
- Maity, P., Hansda, D., Bandyopadhyay, U. and Mishra, D.K. "Biological activities of crude extracts of chemical constituents of Bael, Aegle marmelos (L.) Corr." Indian Journal of Experimental Biology, 2009; 47:849-861.
- Markin, D., Duek, L. and Berdicevsky, I. In vitro antimicrobial activity of olive leaves. Mycoses 2003; 46:132-136.
- Micol, V., Caturla, N., Pérez-Fons, L., Más, V., Pérez, L and Estepa, A. The olive leaf extract exhibits antiviral activity against viral haemorrhagic septicaemia rhabdovirus (VHSV). Antiviral Res. 2005; 66:129-136.
- Motaal, A. A. and Shaker, S. Anticancer and antioxidant activities of standardized whole fruit, pulp, and peel extracts of Egyptian pomegranate. In: The Open Conference Proceedings Journal. 2011; 2:41-45.
- 81. Muthuraman, A., Sood, S. and Singla, S.K. The antiinflammatory potential of phenolic compounds from Emblica officinalis L. in rat. Inflammopharmacology. 2010 Jul 2. [Epub ahead of print]
- 82. Naik, G.H.; Priyadarsini, K. I.and Mohan, H. Evaluating the antioxidant activity of different plant extracts and herbal formulations. Res. Chem. Intermed. 2005; 31(1-3):145-151.
- 83. Negi, P. S.; Jayaprakasha, G. K. and Jena, B.S. Antioxidant and antimutagenic activities of pomegranate peel extracts. Food Chemistry. 2002; 80:393-7.
- 84. Ngamkitidechakul, C., Jaijoy, K., Hansakul, P., Soonthornchareonnon, N. and Sireeratawong, S. Antitumour effects of Phyllanthus emblica induction of cancer cell apoptosis and inhibition of in vivo tumour promotion and in vitro invasion of human cancer cells. Phytother Res, 2010; 24:1405-1413.
- 85. Nicolis, E.; Lampronti, I.and Dechecchi, M.C. Pyrogallol, an active compound from the medicinal plant Emblica officinalis, regulates expression of pro-inflammatory genes in bronchial epithelial cells. Int Immunopharmacol. 2008; 8(12):1672-1680.
- 86. Owen, R.W., Haubner, R., Wurtele, G., Hull, E.,

- Spiegelhalder, B. and Bartsch, H. Olives and olive oil in cancer prevention. Eur J Cancer Prev, 2004; 13:319-326.
- 87. Panichayupakarananta, P.; Issuriya, A.; Sirikatitham, A. and Wang, W. Antioxidant assay-guided purification and LC determination of ellagic acid in pomegranate peel. Journal of Chromatography and Science. 2010; 48(6):456-9.
- 88. Pereira, A.P., Ferreira, I.C., Marcelino, F., Valentao, P., Andrade, P.B., Scabra, R., Estevinho, L., Bento, A and Pereira, J.A. Phenolic compounds and antimicrobial activity of olive (Olea Europea L.ev.Cobrancosa) leaves.Molecules 2007; 12:1153-1162.
- 89. Pieroni, A., Heimler, D., Pieters, L., Van Poel, B and Vlietinck, A.J. In vitro anti-complementary activity of flavonoids from olive (Olea europaea L.) leaves. Pharmazie 1996; 51:765.
- 90. Prior, R. L.; Cao, G.; Martin, A.; Sofic, E.; McEwen, J. and O'Brien, C.; Lischner, N.; Ehlenfeldt, M.; Kalt, W.; Krewer, G. and Mainland, C.M. Antioxidant capacity as influenced by total phenolic and anthocyanin content, maturity, and variety of Vaccinium species. Journal Agric. Food Chem. 1998; 46: 2686-2693.
- 91. Purohit, S.S. and Vyas, S.P. "In: Aegle marmelos Correa ex Roxb,(Bael), Medicinal plant cultivation-A scientific approach. Agrobios, Jodhpur, 2004; 498-504.
- 92. Rajeshkumar, N. V.; Radahakrishna Pillai, M.and Kuttan, R. Inductionof Apoptosis in Mouse and Human Carcinoma Cell Lines by Emblica officinalis Polyphenols and its Effect on Chemical Carcinogenesis. J Exp. Clin. Cancer Res. 2003; 22: 201 212.
- 93. Rao, T.P.; Sakaguchi, N.; Juneja, L.R.; Wada. E. and Yokozawa, T. Amla (Emblica officinalis Gaertn.) extracts reduce oxidative stress in streptozotocin-induced diabetic rats. J Med Food. 2005; 8(3):362-368.
- 94. Rastogi, R.P. and Mehrotra, B.N. Compendium of Indian Plants. CDRI, Lucknow and Publications & Information Directorate, New Delhi. 1993.
- 95. Rice-Evans, C. A. and Miller, N.J. Antioxidant acitivities of flavonoids as bioactive components of food. Biochem. Soc. Trans. 1996; 24:790-795.
- 96. Risch, HA., Jain, M., Marrettt, LD and Howe, GR. 1994. Dietary fat intake and risk of epithelial ovarian cancer. J Natl Cancer Inst 1994; 86:1409–1415.
- 97. Sancheti, G., Jindal, A., Kumari, R. and Goyal. Chemopreventive action of Emblica Officinalis on a skin carcinogenesis in mice. Asian Pacific J Cancer Prev, 2005; 6:197–201.
- 98. Sarwat, S.; Ahmed, S. and Jahangir, T. Emblica officinalis and hepatocarcinogenesis: A chemopreventive study in Wistar rats. J

- Ethnopharmacol. 2008; 118:1-6.
- 99. Sass-Kiss, A. Differences in anthocyanin and carotenoid conent of fruits and vegetables. Food Res Int. 2005; 38:1023-9.
- 100. Saswati Parichha. "Bael (Aegle Marmelos): Nature's Most Natural Medicinal Fruit", Orissa Review. 2004/
- 101. Scartezzini P, Speroni E. Review on some plants of Indian traditional medicine with antioxidant activity. J Ethnopharmacol. 200; 71(1-2):23-43.
- 102. Seeram, N. P.; Aronson, W. J.; Zhang, Y.; Henning, S. M.; Moro, A.; Lee, R. P.; Sartippour, M.; Harris, D. M.; Rettig, M.; Suchard, M. A.; Pantuck, A. J.; Belldegrun, A. and Heber, D. Pomegranate ellagitannin-derived metabolites inhibit prostate cancer growth and localize to the mouse prostate gland. Journal of Agriculture Food Chemistry. 2007; 55:7732-7
- 103. Seeram, N. P.; Lee, R. and Heber, D. Bioavailability of ellagic acid in human plasma after consumption of ellagitannins from pomegranate (Punica granatum L.) juice. Clinical Chim. Acta. 2004; 348:63-8
- 104. Sharma, M.; Li, L.; Celver, J.; Killian, C.; Kovoor, A. and Seeram, N. P. 2010. Effects of fruit ellagitannin extracts, ellagic acid, and their colonic metabolite, urolithin A, on Wnt Signalling. Journal of Agricultural Food Chemistry. 2010; 58(7):3965-9.
- 105. Skrget, M., Kotnik, P., Hadolin, M., Hra, A. R., Simoni, M and Knez, Z. Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. Food Chem. 2005; 89:191-198.
- 106. Somova, L. I., Shode, F. O and Mipando, M. Cardiotonic and antidysrhythmic effects of oleanolic and ursolic acids, methyl maslinate and uvaol. Phytomedicine. 2004; 11:121-129.
- 107. Somova, L. I., Shode, F. O., Ramnanan, P and Nadar, A. Antihypertensive, antiatherosclerotic and antioxidant activity of triterpenoids isolated from Olea europaea, subspecies Africana leaves. J. Ethnopharmacol. 2003; 84:299-30.

- 108. Swami, S., Thakor, M., Patil, M. and Haldankar, P. Jamun (Syzygium cumini L.): A Review of its food and medicinal uses. Food and Nutrition Science. 2012; 3(8):1100-1117.
- 109. Thakur, R.S., Puri, H.S. and Husain, A. Major Medicinal Plants of India. Central Institute of Medicinal and Aromatic Plants, Lucknow, India. 1989.
- 110. Verlangieri, A. J.; Kapeghian, J. C.; el-Dean, S. and Bush, M. Fruit and vegetable consumption and cardiovascular mortality. Med. Hypoth. 1985; 16:7-15.
- 111. Wang, L., Geng, C., Jiang, L., Gong, G., Liu, D., Yoshimura, H and Zhong, L. The anti atherosclerotic effect of olive leaf extract is related to suppressed inflammatory response in rabbits with experimental atherosclerosis. Eur. J. Nut. 2008; 47:235-243.
- 112. Wang, S. Y. and Lin, H. S. Antioxidant activity in fruit and leaves of blackberry, raspberry, and strawberry varies with cultivar and developmental stage. Journal Agri Food Chem. 2000; 48:140-146.
- 113. Wang, C. J.; Wang, J. M.; Lin, W. L.; Chu, C. Y.; Chou, F.P. and Tseng, T.H. Protective effect of Hibiscus anthocyanins against tertbutyl hydeoperoxide-induced hepatic toxicity in rats. Food Chemistry Toxicol. 2002; 38:411-6.
- 114. Willet, C.W. Micronutrients and cancer risk. American Journal Clinical Nutr. 1994; 59:162S-165S.
- 115. Williamson, E.M. Major Herbs of Ayurveda. Churchill- Livingstone, London. 2002.
- 116. Zadernowski, R.; Naczk, M. and Nesterowicz, J. Phenolic acid profiles in some small berries. Journal Agric Food Chem. 2005; 53:2118-24.
- 117. Zhang, L.Z., Zhao, W.H., Guo, Y.J., Tu GZ, Lin S, Xin LG. Studies onchemical constituents in fruits of Tibetan medicine Phyllanthus emblica. Zhongguo Zhong Yao Za Zhi 2003; 28:940 943.